# Nematodes from northern Australian reptiles

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## Abstract

Twenty-one species of nematode were identified from the gastrointestinal tract or body cavity of 131 individual lizards and snakes comprising 26 species. In 13 host species, only a single individual specimen was available, and the aquatic Keelback snake (*Tropidonophis mairii*) comprised almost half the specimens examined. In general, nematode intensity was low, and only a single adult nematode specimen was present in most of the reptiles examined. Most nematode species showed specificity to one host family, except for *Abbreviata bancrofti,* which was recovered from six species across four families. A total of 21 new host-parasite records and one new locality record are listed, with a number of reptile species reported as a host for a nematode for the first time.

## Introduction

Nematode parasites are widespread in snakes and lizards in Australia (Pichelin *et al.* 1999). Gastrointestinal nematodes of varanid lizards and of pythons have been relatively well studied from northern Australia, revealing that these reptiles support a rich nematode fauna, some species showing marked host specificity and geographical range limitations (Jones 1979, 1988; Mulder & Smales 2015). A recent study on the nematodes of the Keelback snake (*Tropidonophis mairii*) showed very high infection prevalence with the spirurid nematode *Tanqua anomala* but with limited negative impacts on the health of the host (Mayer *et al.* 2015). Little is known, however, of nematodes found in other reptile groups in northern Australia. During the course of various projects undertaken on reptiles in this area over many years, reptiles were dissected, and any helminths found were removed for subsequent study. For this paper we examined previously unstudied nematodes dissected from host specimens in the collections of the Museum and Art Gallery of the Northern Territory, the Queensland Museum, and the Australian Museum, as well as from the personal collections of the authors. These nematodes were identified, host specificity and associations between species investigated, and the host habitat and geographical range noted.

## Methods

Preservation of nematode specimens varied, depending on host source (dissections of fresh road-killed specimens or previously preserved museum specimens) and the collector of the nematodes. Thus, some specimens were initially preserved in 10% formaldehyde whereas others were preserved in 70% ethanol; all specimens were subsequently stored in 70% ethanol. Before examination, the nematodes were cleaned then cleared in chlorolactophenol and examined under a BA series Olympus microscope.

Museum reference numbers and infections of every individual reptile examined in this study are given in Table 1. The listed intensity of infection data needs to be treated with caution, especially for records from museum host specimens. Many of these specimens were difficult to dissect due to preservation shape (for example, snake specimens are coiled into jars), the inability to complete a full dissection to preserve the host specimen intact as much as possible, and in many cases, prior dietary studies conducted on the specimens had removed or emptied the contents of their stomach. Accurate records for uninfected snakes sourced from other studies were unavailable, thus prevalence data presented here are incomplete and should also be treated with caution.

## Results

A total of 131 individual host specimens, comprising 26 species across 9 families were found to be infected with nematodes (Table 1). The hosts were lizards of the families Agamidae (3 species), Carphodactylidae (1), Diplodactylidae (2), Gekkonidae (1), Scincidae (2) and Varanidae (3), and snakes of the families Boidae (4), Colubridae (2) and Elapidae (8). Twenty-two reptile species were represented by five or less individuals. The samples were dominated by the aquatic colubrid snake *Tropidonophis mairii*, which accounted for almost half of the host specimens examined.

### *Species identified*

At least 21 nematodes (excluding larvae) were identified from the reptiles examined in this study (Table 2); 13 were identified to species, 6 were identified to genus and 2 could only be identified to family level. Larval and immature stages were also reported (Table 1). Eleven new host-parasite records were observed: *Physalopteroides filicauda* in the Eastern Water Skink (*Eulamprus quoyii*)*; Abbreviata hastaspicula* in the Woma (*Aspidites ramsayi*); *Tropidonophis mairii* in Spencer's Monitor (*Varanus spenceri*); *Abbreviata bancrofti* in *Tropidonophis mairii*, the Lesser Black Whipsnake (*Demansia vestigiata*) and the Eastern Brown Snake (*Pseudonaja textilis*)*; Tanqua tiara* in the Northern Death Adder (*Acanthophis praelongus*); *Dracunculus mulbus* in *Tropidonophis mairii* and the Black-headed Python (*Aspidites melanocephalus*); and *Maxvachonia brygooi* in the Asian House Gecko (*Hemidactylus frenatus*). In addition, the following genera were recorded for the first time from these hosts: *Ophidascaris* sp. 1 in the Coastal Taipan (*Oxyuranus scutellatus*)*; Ophidascaris* sp. in the Orange-Naped Snake (*Furina ornata*); *Pharyngodon* sp. in the Chameleon Gecko (*Carphodactylus laevis*); *Skrjabinodon* sp. and Pharyngodonidae sp. in the Northern Velvet Gecko (*Oedura castelanui*); and *Kalicephalus* sp. in the Slatey-grey Snake (*Stegonotus cucllatus*),

*Tropidonophis mairii* and *Acanthophis praelongus*. The presence of immature *Eustrongylides* sp. in *Tropidonophis mairii* and larval Physalopteridae in the Zigzag Velvet Gecko (*Amalosia rhombifer*) and the Eastern Barred Wedgesnout Ctenotus (*Ctenotus strauchii varius*) are new host records. The report of *Parapharyngodon maplestoni* in *Hemidactylus frenatus* is a new locality record.

*Intensity of infection*

For most of the nematode species, the intensity of infection was low. Additionally, for a large number of records, only one individual of each host species was examined. Thus the true range of intensity of infection remains unknown. In skinks and geckoes, all of which were physically small, the intensity of infection, primarily by pharyngodonids, was generally less than five individuals. However, in snakes, intensity of infection was often greater than 10 individuals, although where more than one species of nematode was present, one was usually present in much higher levels than the other. The parasites with highest intensity were: *Tanqua tiara* in the Yellow-spotted Monitor (*Varanus panoptes*) (intensity over 400); *Abbreviata hastaspicula* in *Varanus spenceri*) (Spencer's Monitor) (intensity of 250) and the Sand Monitor (*Varanus gouldii*) (intensity ranging from 35 to more than 100); and *Abbreviata bancrofti* from *Demansia vestigiata* at a mean intensity of 48 (27–69) for the two Northern Territory specimens and at 17 (1–50) for the six northern Queensland specimens. However, for all of these host species, less than 10 specimens and often only one specimen, were examined. For the most represented host in this study, *Tropidonophis mairii*, the nematode *Tanqua anomala* was recovered at a mean intensity of 31 (1–201). This would be a truer reflection of the range of intensity of infection in this host species as the snakes were specifically examined for infection levels as part of the study by Mayer *et al.* (2015).

*Host specificity*

Pharyngodonid nematodes and physalopterid larvae occurred only in the smaller skinks and geckoes. *Maxvachonia brygooi* was only found in the introduced gecko *Hemidactylus frenatus*. *Strongyluris paronai* only occurred in the agamid lizards. *Kalicephalus* sp., Spiruridae sp., *Eustrongylides* sp. and adults of *Ophidascaris* spp. were all reported only from snakes. *Tanqua anomala* was reported only from *Tropidonophis mairii*, whereas *Tanqua tiara* was reported from a range of snakes and varanids. The various species of *Abbreviata* were found to infect a range of hosts across the major families; for example *Abbreviata bancrofti* was recovered from six species of reptile, across four families.

## Discussion

Most reports of nematode infections in reptiles in Australia are anecdotes associated with the descriptions of new species of nematodes. However, a few recent studies have reported aspects of the ecological relationship between nematodes and their hosts (Barton 2015; Mayer *et al.* 2015; Mulder & Smales 2015). Although a wide range of hosts were examined in this study, just one species, *Tropidonophis mairii*, dominated the study and it is the only host for which good ecological data can be presented.

It is unfortunate that many of the specimens of nematodes collected in this study were not in an optimal condition, being poorly preserved, incomplete, immature, or having only one sex available. And in many cases, only a single individual nematode was recovered. For this reason, full identification to species level was often not possible. However, this study has reported at least nine new host-parasite records and one new locality record, as well as increasing the knowledge of nematodes of many species of reptiles in northern Australia. This is especially true for the Northern Territory, an area that has been severely under-represented in previous studies. *Carphodactylus laevis, Oedura castelnaui, Amalosia rhombifer, Ctenotus strauchii varius, Stegonotus cucullatus, Acanthophis praelongus, Furina ornata, Demansia vestigiata* and *Oxyuranus scutellatus* are all reported as hosts for a nematode parasite for the first time.

In this study, representatives of the nematode genus *Ophidascaris* were recovered from a number of snakes as well as from a single varanid lizard. As discussed above, many of the specimens collected could only been identified to the genus *Ophidascaris*. Both *Ophidascaris moreliae* and *Ophidascaris robertsi* have previously been identified in the Children's Python (*Antaresia childreni*), *Aspidites melanocephalus* and the Carpet Python (*Morelia spilota*), with its congener *Ophidascaris robertsi* reported only from *Aspidites ramsayi* (Mawson 1955; Sprent & Mines 1960; Sprent 1969; Jones 1979). Differentiation of these two closely-related species depends on several characters, including the form of the lips, the presence or absence of a post-oesophageal caecum, cervical alae, and pitting on the surface of the eggs. Identification of specimens to *Ophidascaris moreliae/robertsi* was only possible for nematodes collected from a single *Morelia spilota* from an unknown collection location in northern Queensland. However, the quality of the specimens did not allow for identification to one or the other of the species.

*Ophidascaris pyrrhus* was originally described from a Red-bellied Black Snake (*Pseudechis porphyriacus*) originating from the central New South Wales coast (Johnston & Mawson 1942). It has subsequently been reported from a number of other elapid snakes from across Australia (Jones 1980; Pichelin *et al.* 1999). *Ophidascaris pyrrhus* was only found in one snake (a specimen of *Pseudechis porphyriacus* from northern Queensland) in the present study as an encysted larval stage.

Two males and one female of a species of *Ophidascaris* were recovered from *Oxyuranus scutellatus* from northern Queensland, in which the male copulatory spicules were abnormally long, being more than 10 mm in length, which is twice the length of those reported in *Ophidascaris pyrrhus* (Johnston & Mawson 1942). This suggests that the specimens collected belong to a different, possibly new, species of *Ophidascaris*. However these specimens were not in a sufficiently good condition to compare other morphological features. Nematodes are commonly found in *Oxyuranus scutellatus* (Hoser 2008), but none has previously been identified to species. Further collections of *Oxyuranus scutellatus* from northern Queensland, as well as other locations, are required for the collection of more nematodes to allow for accurate species identification.

The life cycle for most members of the genus *Ophidascaris* remains unknown, however, a mammalian intermediate host is required for development of *Ophidascaris moreliae* to an infective third stage larva (Sprent 1969), whereas *Ophidascaris pyrrhus* is thought to use lizards as its intermediate host (Sprent 1988). The diet of the snakes *Morelia spilota* and *Oxyuranus scutellatus* are both dominated by mammals, whereas the snakes *Antaresia childreni, Aspidites ramsayi,* the Rough-scaled Snake (*Tropidechis carinatus*) and *Pseudonaja textilis* have a more varied diet, with 30–50% being mammals, and *Pseudechis porphyriacus* has a diet dominated by reptiles and amphibians (Shine 1991). Interestingly, none of the colubrid snakes examined in this study were infected with any *Ophidascaris* species. The diet of *Tropidonophis mairii* is almost exclusively amphibians, whereas *Stegonotus cucullatus* has a reptile-dominated diet (Shine 1991), but as only two *Stegonotus cucullatus* were examined in this study, this needs to be treated with caution. *Strongyluris paronai* appears to be restricted to members of the Agamidae (Pichelin *et al.* 1999), and it was reported from both the Frill-neck Lizard (*Chlamydosaurus kingii*) and a Bearded Dragon (*Pogona* sp.) in this study. Previous reports of *Strongyluris paronai* are mainly from the Kimberley region in northern Western Australia (Jones 1986, 1994), with one study of the seasonality of infection in *Chlamydosaurus kingii* in Kakadu National Park (Griffiths *et al.* 1998). Due to the patterns of observation of larval and adult nematodes in *Chlamydosaurus kingii* by Griffiths *et al.* (1998), it is assumed that *Strongyluris paronai* has a direct life cycle, although Anderson (2000) suggested that insects might be utilised as transport hosts, with no development of the parasite within. Both *Chlamydosaurus kingii* and *Pogona* species are known to feed extensively on insects (Cogger 2014).

Representatives of the genus *Maxvachonia* have been reported from a number of lizards, mostly agamids, in Australia, with one dubious report from an elapid snake (Mawson 1972). Species of *Maxvachonia* have also been reported from a few skinks and varanids (Pichelin *et al.* 1999). Further examination of the specimens reported in Barton (2015) collected from *Hemidactylus frenatus* allowed for the specific identification to *Maxvachonia brygooi*. This represents a new host record for *Maxvachonia brygooi*. The life cycle of *Maxvachonia* has not been studied, but other members of the family Cosmocercidae have a direct life cycle, with the infective larva penetrating the host via the mouth or skin (Anderson 2000). As with *Strongyluris paronai*, described above, this nematode may utilise insects as transport hosts, although this has yet to be shown.

Members of the Pharyngodonidae are strictly monoxenous, with direct transmission through the ingestion of infective larvae within eggs (Anderson 2000). Members of the Pharyngodonidae are exclusively found in lizards, especially skinks (Pichelin *et al.* 1999). Pharyngodonids were only collected from geckoes in this study, however all other groups of lizards were only represented by one to three specimens. The vast majority of infections were of *Spauligodon hemidactylus* in *Hemidactylus frenatus*, as reported previously in Barton (2015).

*Parapharyngodon maplestoni* has previously been reported from *Hemidactylus frenatus* in various locations outside of Australia (South-East Asia and Oceania; Barton 2015),

but this is the first published record of this parasite in Australia. As with *Spauligodon hemidactyli* (Barton 2015), it is assumed that *Parapharyngodon maplestoni* has been introduced to Australia with its host. Further research will be required to determine if this species has spread to other native Australian reptiles.

A number of *Pharyngodon* species have been reported from a range of skinks and geckoes across Australia (Pichelin *et al.* 1999). The presence of a species of *Pharyngodon* in *Carphodactylus laevis* is a new host record.

A small number (seven) of species of *Skrjabinodon* have been reported from skinks and geckoes in Australia (Pichelin *et al.* 1999; Jones 2013). The presence of a species of *Skrjabinodon* in *Oedura castelnaui* is a new host record.

*Dracunculus mulbus* was originally reported from the tissues surrounding the organs in the body cavity of the Water Python (*Liasis fuscus*) from Fogg Dam, Northern Territory (Jones & Mulder 2007). *Dracunculus mulbus* occurs in the tissues or body cavity of its host and the true intensity of infection as well as potential range of host species may have been higher. Both *Tropidonophis mairii* and *Aspidites melanocephalus* are new host records, with both species collected from close to the type locality.

It is assumed that *Dracunculus mulbus* has an aquatic-based life cycle, as other members of the genus release larvae into the water from a blister in the skin, and the larvae develop to the infective stage within copepods before being ingested by a paratenic host, such as a tadpole (Anderson 2000). Thus, the aquatic snakes here could be infected via infected frogs, with *Tropidonophis mairi* known to feed primarily on them, although *Liasis fuscus* and *Aspidites melanocephalus* are not known to eat them, preferring either mammals or reptiles (Shine 1991).

The two species of *Tanqua* identified in this collection, *Tanqua tiara* and *Tanqua anomala,* were described by Linstow (1879, 1904) and both were redescribed by Baylis (1916), the former from species of *Varanus* and other aquatic and semi-aquatic reptiles in the Old World tropics, and the latter from the Checkered Keelback (*Xenochrophis piscator*) (previously known as *Tropidonophis piscator*) in Sri Lanka. *Tanqua tiara* occurred in high numbers in the single specimen of *Varanus panoptes* examined, and has been recorded at moderately high prevalence (25% to greater than 50%), at intensities of up to 120, in three species of *Varanus* from northern Australia (Jones 1988). *Tanqua anomala* occurred exclusively in the aquatic colubrid *Tropidonophis mairii*, with a mean intensity of 31 from the snakes examined in this study. Mayer *et al.* (2015) reported a mean intensity of 35 nematodes (0–243) for 93 *Tropidonophis mairii* examined from Fogg Dam; it is from these snakes that the nematodes examined in this study were taken.

The cases of *Tanqua tiara* infections reported in this study were from the Darwin and Fogg Dam region. Additionally, only in the Northern Territory was *Tropidonophis mairii* infected with *Tanqua anomala*, with none of the four *Tropidonophis mairii* collected from Queensland infected with any species of *Tanqua*. The collection location of *Tropidonophis*

*mairii* in Queensland is an area affected by tidal intrusion, thus the required intermediate hosts may not be able to survive in this area; however, further collections of snakes from other areas of northern Queensland are required to determine if this parasite does occur there or is restricted to the Northern Territory.

Physalopterid nematodes are the dominant gastric nematode in a number of Australian reptile families (Jones 1991, 2014). Adults of *Abbreviata* are the dominant genus in the larger reptiles (Jones 1991, 2014).The high intensities of *Abbreviata hastaspicula* in species of *Varanus* have been previously documented by Jones (2014). Physalopterid nematodes possess a two host life cycle, with an arthropod intermediate host and a paratenic host where the larva is found coiled within a cyst in the stomach wall (Jones 1991, 2014), as was found often in this study in the smaller species of reptiles examined. Identification of these larvae to species cannot be done through morphology alone (Jones 1991). It is unknown whether this cyst stage is a prerequisite for final maturity or whether there is some physiological clue in the (usually) larger final host for development to occur to adult (Jones 1991). Adult physalopterids were thought to be absent in colubrid snakes, with the exception of an unpublished report in Jones (2014). Reports from boid snakes were considered spurious, with nematodes found thought to actually have been infections in the prey items (Jones 2014). This study, however, confirms the presence of *Abbreviata* species in members of both these families in Australia.

*Abbreviata bancrofti* has a widespread distribution, and had been reported from 21 species across three families (Jones 2014). The colubrid *Tropidonophis mairii* is confirmed as a host species, as well as the additional elapids *Demansia vestigiata* and *Pseudonaja textilis*. The boids *Aspidites melanocephalus* and *Morelia spilota* are also reported as hosts in this study with infection levels similar to those found in the 'usual' hosts, which would tend to confirm that they are true hosts for these parasites. *Abbreviata bancrofti* is found in regions along the coastal eastern and southern fringe of Australia as well as the northern tropics in areas with higher rainfall levels (Jones 2014).

*Abbreviata hastaspicula* if predominantly found in drier areas and the northern tropics where the mean annual temperature is greater than 18°C (Jones 2014). *Abbreviata hastaspicula* had been exclusively reported from varanid lizards (see Jones 2014); this study reports the snakes *Aspidites ramsayi*, *Stegonotus cucullatus* and *Tropidonophis mairii* as hosts, although at much lower levels of infection that that reported for varanids.

Most of the reports for *Abbreviata* spp. were for immature specimens which were unable to be identified to species due to the lack of required taxonomic characters.

*Physalopteroides filicauda* was originally described from the Smooth Knob-tail Gecko (*Nephurus laevissimus*) (Jones 1985) and has subsequently been reported from a variety of lizards across all families, with the majority of records from Western Australia (Pichelin *et al.* 1999; Goldberg & Bursey 2012).

Members of the genus *Kalicephalus* have been reported from a small number of snakes across Australia (Pichelin *et al.* 1999). Unfortunately, none of the specimens collected in this study was suitable for providing a species identification, although they were considered closest to *Kalicephalus australiensis*. The life cycle of *Kalicephalus*, like other strongylids, is considered direct via ingestion of the infective larva, although the use of paratenic hosts cannot be ruled out (Anderson 2000). All of the infected snakes in this study came from Fogg Dam and all are new host records for members of this genus of nematode in Australia.

There are two previous records of *Eustrongylides* in Australia: *Eustrongylides acrochordi* (immature female) from the stomachs of two (out of eight) Arafura File Snakes (*Acrochordus arafurae*) (Jones 1978), an aquatic fish-eating snake, plus encysted in the livers of water pythons (*Liasis fuscus*) (Mulder & Smales 2015). Nematodes in the genus *Eustrongylides* usually have two intermediate hosts, with fishes being the usually recorded second intermediate host (Anderson 1992). Species of *Eustrongylides* mature in the proventriculus of fish-eating aquatic birds, and it is likely that *Acrochordus arafurae*, *Liasis fuscus* and *Tropidonophis mairii* were paratenic or accidental hosts (Jones, 1978; Mulder & Smales 2015; present study).

The host-specificities of nematodes recovered during this study correspond with those of previous studies (Jones 1980, 1983, 1988). The low host specificity of *Abbreviata bancrofti* has been noted previously (Jones 2014). The habitat range of this nematode's hosts is exemplified by its high prevalence and intensity in *Demansia vestigiata*, which normally inhabits drier habitats (Cogger 2014), and it was the second most common nematode in the aquatic *Tropidonophis mairii*. In 16 host species, only a single adult nematode specimen was recovered (Table 2). The intensity of most nematodes was low, and this finding in several host species accords with the conclusions of Mulder & Smales (2015) for infections in the python *Liasis fuscus*. In some cases however the finding of a single nematode may have been the result of accidental infection, the nematode having been ingested within the prey.

This study, therefore, confirms previous observations on host specificities and intensities of infection, and extends the known hosts of several species. Further studies with larger samples would provide a fuller picture of the nematode fauna of reptiles in northern Australia. Such basic data is needed to monitor possible faunal changes with both the advance of the Cane Toad (*Rhinella marina*) and changes in climate.

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Sprent J.F.A. and Mines J.J. (1960) A new species of *Amplicaecum* (Nematoda) from the carpet snake (*Morelia argus variegatus*): with a re-definition and a key for the genus. *Parasitology* 50, 183–198.

**Table 1.** Individual reptile hosts of nematodes examined in this study, separated into A) lizards and B) snakes. Sources of hosts are listed as those dissected from the collections of the Museum and Art Gallery of the Northern Territory (NTM), the Queensland Museum (QM) and the Australian Museum (AM) and from specimens donated to DPB by staff of the University of Sydney Tropical Ecology Research Facility (TERF) or dissected by DPB at James Cook University, Townsville (JCU). The host registration number for museum specimens, where applicable, is listed; registration numbers for the collected nematodes is provided in the last column. Unreg denotes an unregistered specimen; unk denotes unknown information.

**A. LIZARDS**

| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| Agamidae | *Chlamydosaurus kingii* | NT | Elcho Island | JCU | *Strongyluris paronai* | Adult | Stomach | 1 | NTM D1810 |
| | *Intellagama lesueurii* | Qld | Tully Rd/Landstone Rd, Tully | JCU | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1778 |
| | *Pogona* sp. | Qld | No collection data | JCU | *Strongyluris paronai* | Adult | Intestine | 6 | NTM D1809 |
| Carphodactylidae | *Carphodactylus laevis* | Qld | Iron Range | JCU | *Pharyngodon* sp. | Adult | Intestine | 2 | N/A |
| Diplodactylidae | *Amalosia rhombifer* | NT | Cape Fourcroy, Bathurst Island | NTM R7889 | Physalopteridae sp. | Larva | Wall of Stomach | 1 | NTM D1812 |
| | | | | NTM R7969 | Physalopteridae sp. | Larva | Wall of Stomach | 1 | N/A |
| | *Oedura castelnaui* | Qld | Ayr District | JCU | Pharyngodonidae sp. | Adult | Rectum | 1 | NTM D1828 |
| | | | | | *Skrjabinodon* sp. | Adult | Rectum | 1 | NTM D1798 |
| Gekkonidae | *Hemidactylus frenatus* | NT | Beatrice Hill | NTM R1640 | *Spauligodon hemidactylus* | Adult | Rectum | 3 | NTM D1648 |
| | | | Berry Springs Reserve | NTM R6346 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1631 |
| | | | Black Point | NTM R20939 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1639 |
| | | | Croker Is, Mindjilang Community | NTM R20409 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | N/A |
| | | | Darwin | NTM R19816 | *Maxvachonia brygooi* | Adult | Intestine | 1 | NTM D1819 |
| | | | | | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1638 |
| | | | | NTM R25671 | *Parapharyngodon maplestoni* | Adult | Rectum | 5 | NTM D1641 |
| | | | | NTM R25980 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1822 |
| | | | | NTM R0010 | *Spauligodon hemidactylus* | Adult | Rectum | 10 | NTM D1625 |
| | | | | NTM R19815 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1637 |
| | | | Katherine | NTM R8142 | *Spauligodon hemidactylus* | Adult | Intestine | 1 | NTM D1632 |
| | | | New Year Island | NTM R7687 | Physalopteridae sp. | Larva | Wall of Stomach | 1 | N/A |
| | | | Ti Tree | NTM R6564 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1829 |
| | | WA | Christmas Island | NTM R19503 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | N/A |
| | | Unknown | No collection data | NTM R3566 | *Spauligodon hemidactylus* | Adult | Rectum | 1 | NTM D1628 |
| Scincidae | *Ctenotus strauchii varius* | NSW | Sturt National Park | AM R151609 | Physalopteridae sp. | Larva | Intestine | 1 | AM W49136 |
| | | | | AM R152920 | Physalopteridae sp. | Larva | Intestine | 1 | AM W49137 |
| | | | | AM R152957 | Physalopteridae sp. | Larva | Intestine | 1 | AM W49138 |

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| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| | *Eulamprus quoyii* | Qld | Mimosa Creek, Blackdown Tableland, near Rockhampton | JCU | *Physalopteroides filicauda* | Adult | Intestine | 1 | NTM D1827 |
| Varanidae | *Varanus gouldii* | NT | Darwin | NTM unreg | *Abbreviata hastaspicula* | Adult | Stomach | 105 | NTM D1771 |
| | | | | NTM unreg | *Abbreviata bancrofti* | Adult | Stomach | 9 | NTM D1747 |
| | | | | | *Abbreviata hastaspicula* | Adult | Stomach | 51 | NTM D1747 |
| | | | | | *Ophidascaris* sp. | Larva | Stomach | 1 | NTM D1797 |
| | | | | | *Tanqua tiara* | Adult | Stomach | 2 | NTM D1782 |
| | | | | NTM unreg | *Abbreviata bancrofti* | Adult | Stomach | 10 | NTM D1772 |
| | | | | | *Abbreviata hastaspicula* | Adult | Stomach | 35 | NTM D1772 |
| | | | | | *Tanqua tiara* | Adult | Stomach | 1 | NTM D1781 |
| | | Qld | Ayr | NTM unreg | *Abbreviata* spp. | Adult | Stomach | 1 | NTM D1775 |
| | *Varanus panoptes* | NT | Darwin | NTM unreg | *Tanqua tiara* | Adult | Stomach | 420 | NTM D1786 |
| | *Varanus spenceri* | NT | Barkly Tablelands | NTM R28049 | *Abbreviata hastaspicula* | Adult | Stomach | 250 | NTM D1774 |
| **B. SNAKES** | | | | | | | | | |
| **Host Family** | **Host Species** | **State Collected** | **Location Collected** | **Source** | **Identification** | **Stage** | **Location in host** | **No.** | **Parasite Museum Deposit Number** |
| Boidae | *Antaresia childreni* | NT | Fogg Dam | TERF | *Ophidascaris* spp. | Adult | Intestine | 25 | NTM D1807 |
| | *Aspidites melanocephalus* | NT | Darwin | NTM R8254 | *Dracunculus mulbus* | Larva | Mesenteries | 1 | NTM D1633 |
| | | | Katherine | NTM R242 | *Abbreviata bancrofti* | Adult | Stomach | 6 | NTM D1626 |
| | | Qld | Clermont-Alpha Road | QM J60006 | *Abbreviata bancrofti* | Adult | Intestine | 8 | QM G222593 |
| | | | Longreach-Jundah Hwy, near Stonehenge | QM J73843 | *Abbreviata bancrofti* | Adult | Intestine | 9 | QM G222591 |
| | | Unknown | No collection data | QMJ2259 | *Abbreviata* spp. | Adult | Intestine | 1 | QM G222590 |
| | *Aspidites ramsayi* | NT | Frewana | NTM R8498 | *Abbreviata hastaspicula* | Adult | Stomach | 5 | NTM D1634 |
| | | | | | *Ophidascaris* spp. | Larva | Body wall | 1 | NTM D1806 |
| | | | No collection data | NTM unreg | *Ophidascaris* spp. | Adult | Stomach | 5 | NTM D1706 |
| | | | Stirling Creek | NTM R31612 | *Abbreviata hastaspicula* | Adult | Stomach | 26 | NTM D1643 |
| | | | | | *Abbreviata* spp. | Adult | Rectum | 1 | NTM D1780 |
| | | | Stuart Highway, 116km E of Three Ways | QM J52856 | *Abbreviata hastaspicula* | Adult | Intestine | 10 | QM G222595 |
| | | | Tennant Creek | NTM R26517 | *Abbreviata hastaspicula* | Adult | Intestine | 1 | NTM D1642 |
| | | Qld | Bedourie | QM J18010 | *Ophidascaris* spp. | Adult | Stomach | 1 | QM G222596 |
| | | | Glenmorgan area | QM J40053 | *Abbreviata* spp. | Adult | Intestine | 3 | QM G222594 |
| | *Morelia spilota* | Qld | Bruce Highway, Ayr | JCU | *Ophidascaris* spp. | Adult | Stomach | 1 | NTM D1805 |
| | | | Kareesa Power station, Cardstone Rd | JCU | *Ophidascaris* spp. | Adult | Stomach | 1 | NTM D1808 |

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| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| | | | Nelly Bay, Magnetic Island | JCU | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1776 |
| | | | Tomoulin (near Ravenshoe) | JCU | *Ophidascaris* spp. | Adult | Stomach | 3 | NTM D1802 |
| | | | No collection data | JCU | *Ophidascaris moreliae/ robertsi* | Adult | Stomach | 9 | NTM D1800 |
| Colubridae | *Stegonotus cucullatus* | NT | Fogg Dam | TERF | *Abbreviata hastaspicula* | Adult | Intestine | 1 | NTM D1768 |
| | | | | | *Tanqua tiara* | Adult | Intestine | 1 | N/A |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1745 |
| | | | | | *Kalicephalus* sp. | Adult | Intestine | 1 | NTM D1814 |
| | *Tropidonophis mairii* | NT | Darwin | TERF | *Unknown* | Larva | Mesenteries | 1 | NTM Unreg |
| | | | Fogg Dam | TERF | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1714 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 20 | NTM D1670 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 10 | NTM D1720 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 135 | NTM D1682 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 7 | NTM D1742 |
| | | | | | *Dracunculus mulbus* | Adult | Intestine | 1 | NTM D1815 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 7 | NTM D1707 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 21 | NTM D1722 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 145 | NTM D1687 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 135 | NTM D1705 |
| | | | | TERF | *Eustrongylides* sp. | Larva | Intestine | 1 | NTM D1787 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 45 | NTM D1691 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 2 | NTM D1737 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 12 | NTM D1674 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 4 | NTM D1710 |
| | | | | | *Eustrongylides* sp. | Larva | Intestine | 1 | NTM D1792 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1732 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 22 | NTM D1658 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 32 | NTM D1715 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 21 | NTM D1698 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 3 | NTM D1731 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 54 | NTM D1692 |
| | | | | TERF | *Abbreviata* spp. | Larva | Intestine | 3 | NTM D1736 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 24 | NTM D1659 |
| | | | | TERF | *Kalicephalus* sp. | Adult | Intestine | 1 | NTM D1793 |

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| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| | | | | | *Tanqua anomala* | Adult | Intestine | 4 | NTM D1699 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 8 | NTM D1709 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 60 | NTM D1669 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 6 | N/A |
| | | | | | *Abbreviata* spp. | Adult | Intestine | 5 | NTM D1741 |
| | | | | | *Kalicephalus* sp. | Adult | Intestine | 1 | NTM D1794 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 201 | NTM D1690 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 2 | NTM D1718 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 50 | NTM D1678 |
| | | | | TERF | *Abbreviata hastaspicula* | Adult | Intestine | 1 | NTM D1746 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 37 | NTM D1673 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 2 | NTM D1708 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 2 | NTM D1752 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 3 | NTM D1740 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 54 | NTM D1696 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 17 | NTM D1734 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 26 | NTM D1695 |
| | | | | TERF | *Abbreviata bancrofti* | Larva | Intestine | 1 | NTM D1719 |
| | | | | | *Eustrongylides* sp. | Larva | Intestine | 1 | NTM D1791 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 10 | NTM D1683 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1738 |
| | | | | | *Tanqua anomala* | Unk | Intestine | 4 | NTM D1703 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 54 | NTM D1676 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 8 | NTM D1733 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 56 | NTM D1660 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 17 | NTM D1667 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1743 |
| | | | | | *Eustrongylides* sp. | Larva | Intestine | 4 | NTM D1789 |
| | | | | | *Kalicephalus* sp. | Larva | Intestine | 1 | NTM D1795 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 1 | NTM D1702 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 21 | NTM D1675 |
| | | | | TERF | *Abbreviata hastaspicula* | Adult | Intestine | 1 | NTM D1744 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 10 | NTM D1684 |
| | | | | TERF | *Eustrongylides* sp. | Larva | Intestine | 2 | NTM D1790 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 9 | NTM D1677 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 26 | NTM D1751 |
| | | | | | *Eustrongylides* sp. | Larva | Intestine | 1 | NTM D1788 |

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| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| | | | | | *Tanqua anomala* | Adult | Intestine | 7 | NTM D1784 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 2 | NTM D1713 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 20 | NTM D1701 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 11 | NTM D1766 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 10 | NTM D1783 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 34 | NTM D1749 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 6 | NTM D1721 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 12 | NTM D1689 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 2 | NTM D1729 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 19 | NTM D1688 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1724 |
| | | | | | *Dracunculus* sp. | Adult | Tissues | 1 | NTM D1816 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 20 | NTM D1672 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 19 | NTM D1716 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 15 | NTM D1680 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1717 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 8 | NTM D1668 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 1 | NTM D1735 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 13 | NTM D1661 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 5 | NTM D1711 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 5 | NTM D1704 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 8 | NTM D1727 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 25 | NTM D1693 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 1 | NTM D1666 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 6 | NTM D1657 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 65 | NTM D1706 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 3 | NTM D1748 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 23 | NTM D1785 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 6 | NTM D1750 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 1 | NTM D1761 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 22 | NTM D1726 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 16 | NTM D1686 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 7 | NTM D1730 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 10 | NTM D1656 |
| | | | | TERF | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1739 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 9 | NTM D1663 |
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 10 | NTM D1662 |

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| Host Family | Host Species | State Collected | Location Collected | Source | Identification | Stage | Location in host | No. | Parasite Museum Deposit Number |
|---|---|---|---|---|---|---|---|---|---|
| | | | | TERF | *Tanqua anomala* | Adult | Intestine | 1 | NTM D1681 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 32 | NTM D1728 |
| | | | | | *Tanqua anomala* | Adult | Intestine | 24 | NTM D1679 |
| | | Qld | Cape Cleveland | JCU | *Abbreviata bancrofti* | Adult | Stomach | 4 | NTM D1765 |
| | | | | | *Spiruridea* sp. | Adult | Stomach | 5 | N/A |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 5 | NTM D1759 |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 1 | NTM D1754 |
| | | | | JCU | *Abbreviata* spp. | Adult | Stomach | 4 | NTM D1755 |
| Elapidae | *Acanthophis praelongus* | NT | Fogg Dam | TERF | *Unknown* | Larva | Intestine | 1 | NTM unreg |
| | | | | TERF | *Kalicephalus* sp. | Adult | Intestine | 1 | NTM D1813 |
| | | | | TERF | *Tanqua tiara* | Adult | Intestine | 1 | NTM D1665 |
| | *Furina ornata* | NT | Fogg Dam | TERF | *Ophidascaris* spp. | Adult | Intestine | 41 | NTM D1803 |
| | *Demansia vestigiata* | NT | Fogg Dam | TERF | *Abbreviata bancrofti* | Adult | Intestine | 27 | NTM D1758 |
| | | | | TERF | *Abbreviata bancrofti* | Adult | Intestine | 69 | NTM D1767 |
| | | Qld | Cape Cleveland | JCU | *Abbreviata bancrofti* | Adult | Stomach | 1 | NTM D1773 |
| | | | Townsville | JCU | *Abbreviata bancrofti* | Adult | Stomach | 41 | NTM D1756 |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 4 | NTM D1769 |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 2 | NTM D1760 |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 50 | NTM D1763 |
| | | | | JCU | *Abbreviata bancrofti* | Adult | Stomach | 5 | NTM D1757 |
| | | | Nelly Bay, Magnetic Island | JCU | *Abbreviata* spp. | Adult | Stomach | 2 | NTM D1770 |
| | *Oxyuranus scutellatus* | Qld | Tinaroo Falls | JCU | *Ophidascaris* sp. *1* | Adult | Stomach | 3 | NTM unreg |
| | *Pseudechis australis* | NT | Howard Springs | NTM R5245 | *Abbreviata* spp. | Adult | Intestine | 10 | NTM D1630 |
| | *Pseudechis porphyriacus* | Qld | Paluma Dam | JCU | *Ophidascaris pyrrhus* | Larva | Wall of Stomach | 7 | NTM D1801 |
| | | | | JCU | *Abbreviata* spp. | Adult | Intestine | 1 | NTM D1779 |
| | *Pseudonaja textilis* | NT | Cockatoo Ck crossing | NTM unreg | *Abbreviata bancrofti* | Adult | Stomach | 1 | NTM D1764 |
| | | | | | *Ophidascaris* spp. | Adult | Stomach | 1 | NTM D1796 |
| | | Qld | Bruce Highway, S of Townsville | JCU | *Abbreviata bancrofti* | Adult | Intestine | 27 | NTM D1777 |
| | | | Gootchy Ck turnoff, Gootchy Ck Road, Bruce Highway | JCU | *Ophidascaris* spp. | Adult | Stomach | 1 | NTM D1804 |
| | *Tropidechis carinatus* | NSW | Kingscliffe Beach | QM J21054 | *Ophidascaris* spp. | Adult | Stomach | 1 | QM G222597 |
| | | Qld | Noosa area | QM J25983 | *Ophidascaris* spp. | Adult | Stomach | 1 | |

**Table 2.** List of nematode species and their hosts found in this study. Prevalence is given as the number of hosts infected from the sample of hosts examined. Intensity of infection is presented as the range with the mean in parentheses. The abbreviation Unk denotes unknown information. Symbols: * = New host record; # = New locality record; † = possibly an undescribed species.

| Order | Family | Identification | Host Family | Host Species | State Collected | Prevalence | Intensity |
|---|---|---|---|---|---|---|---|
| Ascaridida | Ascarididae | *Ophidascaris morelaie/robertsi* | Boidae | *Morelia spilota* | Qld | 1/5 | 9 |
| | | *Ophidascaris* sp. 1† | Elapidae | *Oxyuranus scutellatus** | Qld | 1/1 | 3 |
| | | *Ophidascaris pyrrhus* | Elapidae | *Pseudechis porphyriacus* | Qld | 1/1 | 7 |
| | | *Ophidascaris* spp. | Boidae | *Antaresia childreni* | NT | 1/1 | 25 |
| | | | | *Aspidites ramsayi* | NT | 2/5 | 1–5 (3) |
| | | | | | Qld | 1/2 | 1 |
| | | | | *Morelia spilota* | Qld | 3/5 | 1–3 (1.7) |
| | | | Colubridae | *Tropidechis carinatus* | NSW | 1/1 | 1 |
| | | | | | Qld | 1/1 | 1 |
| | | | Elapidae | *Furina ornata** | NT | 1/1 | 41 |
| | | | | *Pseudonaja textilis* | NT | 1/1 | 1 |
| | | | | | Qld | 1/2 | 1 |
| | | | Varanidae | *Varanus gouldii* | NT | 1/3 | 1 |
| | Cosmocercidae | *Maxvachonia brygooi* | Gekkonidae | *Hemidactylus frenatus** | NT | 1/12 | 1 |
| | Heterakidae | *Strongyluris paronai* | Agamidae | *Chlamydosaurus kingii* | NT | 1/1 | 1 |
| | | | | *Pogona* sp. | Qld | 1/1 | 6 |
| Enoplida | Dioctophymatidae | *Eustrongylides* sp. | Colubridae | *Tropidonophis mairii** | NT | 6/54 | 1–4 (1.7) |
| Oxyurida | Pharyngodonidae | *Parapharyngodon maplestoni* | Gekkonidae | *Hemidactylus frenatus#* | NT | 1/12 | 5 |
| | | *Pharyngodon* sp. | Gekkonidae | *Carphodactylus laevis** | Qld | 1/1 | 2 |
| | | *Pharyngodonidae* sp. | Gekkonidae | *Oedura castelnaui** | Qld | 1/1 | 1 |
| | | *Skrjabinodon* sp. | Gekkonidae | *Oedura castelnaui** | Qld | 1/1 | 1 |
| | | *Spauligodon hemidactylus* | Gekkonidae | *Hemidactylus frenatus* | NT | 10/12 | 1–10 (2.3) |
| | | | | | WA | 1/1 | 1 |
| | | | | | Unk | 1/1 | 1 |
| Spirurida | Dracunculidae | *Dracunculus mulhus* | Boidae | *Aspidites melanocephalus** | NT | 1/2 | 1 |
| | | | Colubridae | *Tropidonophis mairii** | NT | 1/54 | 1 |
| | | *Dracunculus* sp. | Colubridae | *Tropidonophis mairii* | NT | 1/54 | 1 |
| | Gnathostomatidae | *Tanqua anomala* | Colubridae | *Tropidonophis mairii* | NT | 51/54 | 1–201 (31.2) |
| | | *Tanqua tiara* | Colubridae | *Stegonotus cucullatus* | NT | 1/2 | 1 |
| | | | Elapidae | *Acanthophis praelongus** | NT | 1/3 | 1 |
| | | | Varanidae | *Varanus gouldii* | NT | 2/3 | 1–2 (1.5) |
| | | | | *Varanus panoptes* | NT | 1/1 | 420 |

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| Order | Family | Identification | Host Family | Host Species | State Collected | Prevalence | Intensity |
|---|---|---|---|---|---|---|---|
| | Physalopteridae | *Abbreviata hancrofti* | Boidae | *Aspidites melanocephalus* | NT | 1/2 | 6 |
| | | | | | Qld | 2/2 | 8–9 (8.5) |
| | | | | *Morelia spilota* | Qld | 1/5 | 1 |
| | | | Colubridae | *Tropidonophis mairii** | NT | 30/54 | 1–32 (8.9) |
| | | | | | Qld | 3/4 | 1–5 (3.3) |
| | | | Elapidae | *Demansia vestigiata** | NT | 2/2 | 27–69 (48) |
| | | | | | Qld | 6/7 | 1–50 (17.2) |
| | | | | *Pseudonaja textilis** | NT | 1/1 | 1 |
| | | | | | Qld | 1/2 | 27 |
| | | | Varanidae | *Varanus gouldii* | NT | 2/3 | 9–10 (9.5) |
| | | *Abbreviata hataspicula* | Boidae | *Aspidites ramsayi** | NT | 4/5 | 1–26 (10.5) |
| | | | Colubridae | *Stegonotus cucullatus* | NT | 1/2 | 1 |
| | | | | *Tropidonophis mairii** | NT | 2/54 | 1 |
| | | | Varanidae | *Varanus gouldii* | NT | 3/3 | 35–105 (63.7) |
| | | | | *Varanus spenceri** | NT | 1/1 | 250 |
| | | *Abbreviata* spp. | Agamidae | *Intellagama lesueurii* | Qld | 1/1 | 1 |
| | | | Boidae | *Aspidites melanocephalus* | Unk | 1/1 | 1 |
| | | | | *Aspidites ramsayi* | NT | 1/5 | 1 |
| | | | | | Qld | 1/2 | 3 |
| | | | Colubridae | *Stegonotus cucullatus* | NT | 1/2 | 1 |
| | | | | *Tropidonophis mairii* | NT | 8/54 | 1–7 (2.9) |
| | | | | | Qld | 1/4 | 4 |
| | | | Elapidae | *Demansia vestigiata* | Qld | 1/7 | 2 |
| | | | | *Pseudechis australis* | NT | 1/1 | 10 |
| | | | | *Pseudechis porphyriacus* | Qld | 1/1 | 1 |
| | | | Varanidae | *Varanus gouldii* | Qld | 1/1 | 1 |
| | | *Physalopteridae* sp. | Gekkonidae | *Hemidactylus frenatus* | NT | 1/12 | 1 |
| | | | | *Oedura rhombifer** | NT | 2/2 | 1 |
| | | | Scincidae | *Ctenotus strauchii varius** | NSW | 3/3 | 1 |
| | | *Physalopteroides filicauda* | Scincidae | *Eulamprus quoyii** | Qld | 1/1 | 1 |
| | Spiruridae | Spiruridae sp. | Colubridae | *Tropidonophis mairii* | Qld | 1/4 | 5 |
| Strongylida | Diaphanocephalidae | *Kalicephalus* sp. | Colubridae | *Stegonotus cucullatus** | NT | 1/2 | 1 |
| | | | | *Tropidonophis mairii** | NT | 3/54 | 1 |
| | | | Elapidae | *Acanthophis praelongus** | NT | 1/3 | 1 |
| Unknown | Unknown | Unknown | Colubridae | *Tropidonophis mairii* | NT | 1/61 | 1 |
| | | | Elapidae | *Acanthophis praelongus* | NT | 1/3 | 1 |